

# Forage Conference at GCA Convention

## Bermudagrass Stem Maggot: Agronomic Research Update

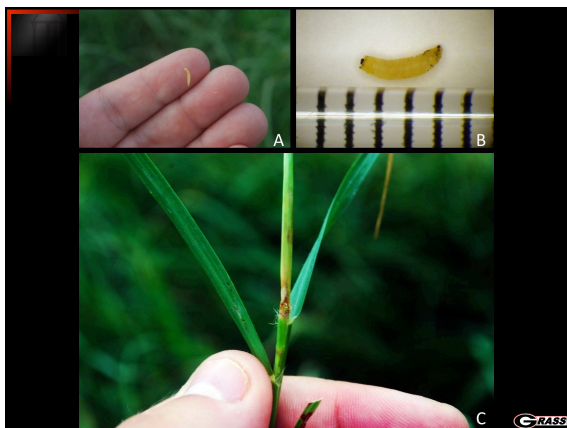
**Bermudagrass Stem Maggot**

Georgia Beef Cattle Short Course  
Feb. 28, 2017

Bill Anderson, Dennis Hancock, Will Hudson







**Stem boring maggot (fly) damaging bermudagrass hay fields, Irwin County, GA, July 2010**



Photos by: Will Hudson, UGA Entomology




**Bermudagrass stem maggot, *Atherigona reversura*, native to Asia**



Photo by: Will Hudson, UGA Entomology




600 μm




Photo by: Lisa Baxter, UGA Crop & Soil Sciences




(Larval photos by Ruth Donaldson, UGA Griffin)

**History**

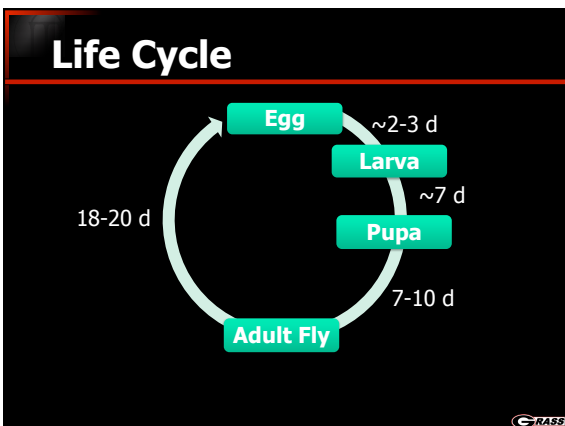
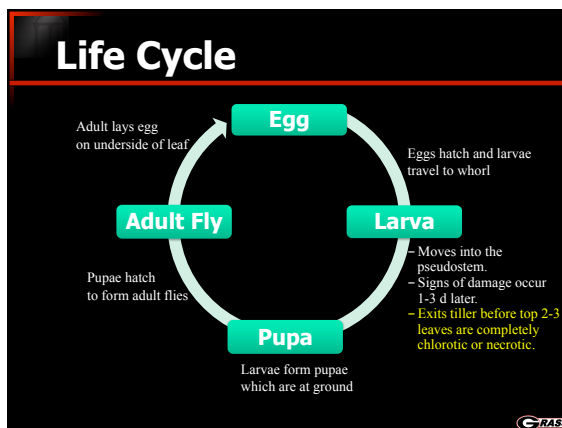
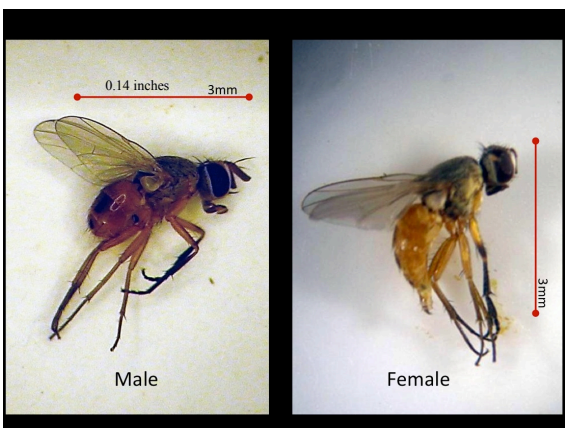
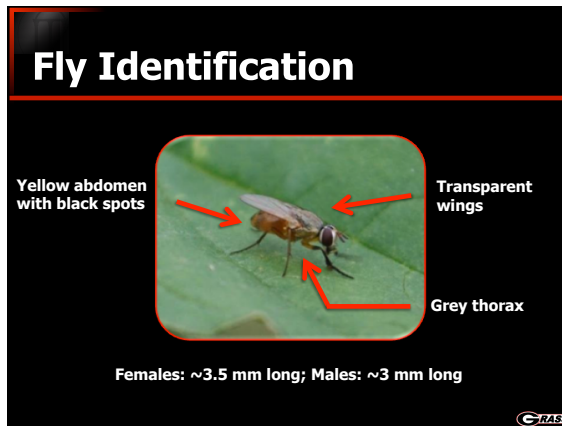
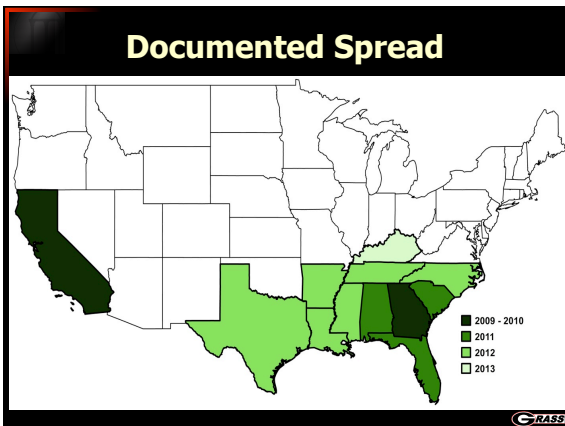
- First recorded (Villeneuve, 1936)
  - Southeast Asia
  - Now found in 20+ countries (Pont et al., 1995)
- January 1974, Hawaii (Hardy, 1976; Hardy, 1981)
- July 2009, California (Holderbaum, 2009)
- Summer 2010, Georgia (Hancock, 2012)
- Now reported throughout southeast



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### Collecting Flies and Larvae

- Sweep nets are quite effective.
  - Flies do not fly far (< 10 ft.)
  - Rarely fly high (< 18 in.)
  - Hard to sweep if dew present.
- Fly populations are high
  - Often 50-80 flies/10 sweeps
  - ~300-500,000 flies/acre

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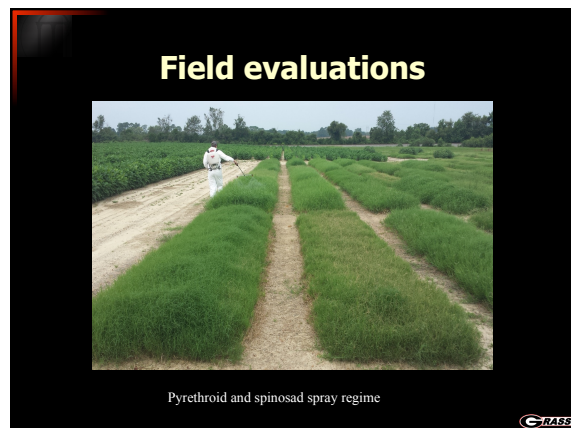
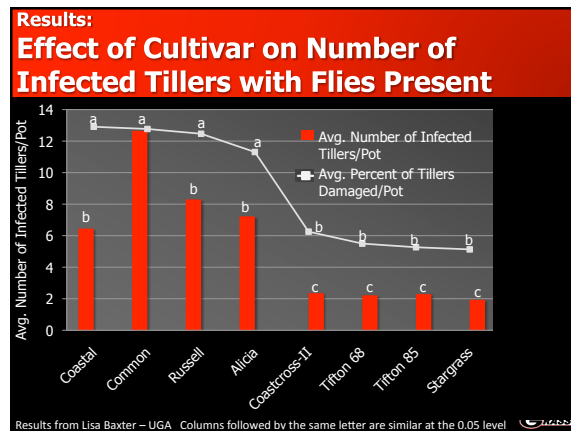
### Chambers to check on flies or fly traps



**GRASS**

### Collecting Larvae

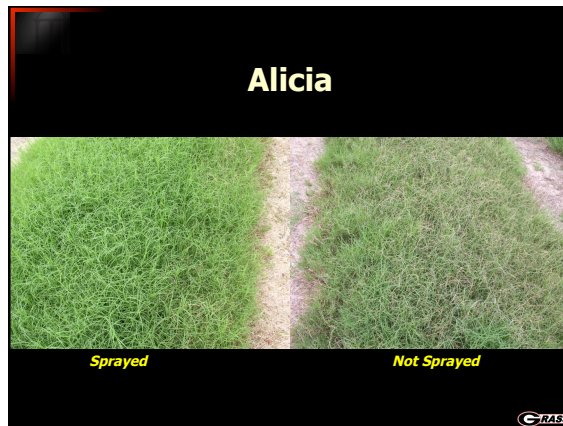
- Dissect pseudostems showing first signs of chlorosis
  - Sharp knife or razor blade
  - Work over dark surface
- No protocols yet for finding pupae in soil.
  - We don't know yet if the pupae overwinter

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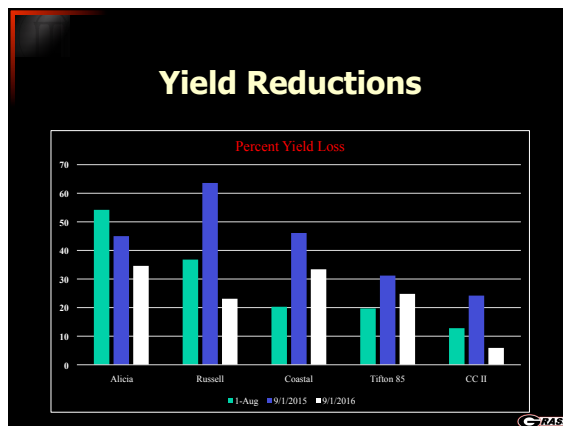
### Screening for Resistance

Rate for BSM      Harvest of core

**Tolerant lines**

Entry	Yield	% Reduction
PI 294467	2263 a	6.6 a
PI 290812	1845 b	9.5 ab
Breeding Line	2350 a	10.3 ab
PI 290664	2170 a	11.1 ab
PI 290872	2104 ab	19.2 ab
Tifton 85	1493 c	18.5 ab
Higgs	1022 d	47.4 c
Alicia	616 e	40.3 c

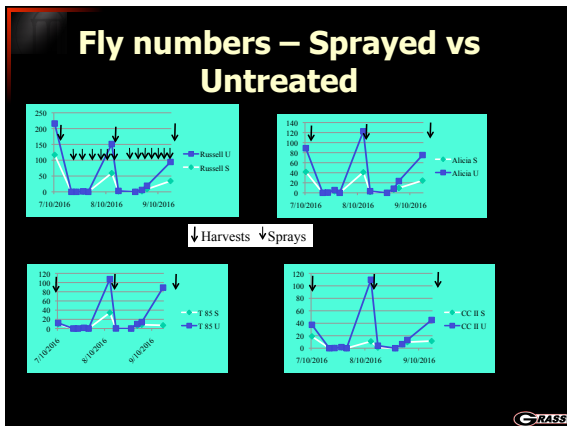
Side-by-side yield evaluations of selected lines



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### Mitigation Strategies

Strategy is contingent upon timing

- If signs of BSM damage occur when the bermudagrass is 6-8 in. tall, (~3 wk after prev. cutting/grazing), then clip (remove if possible) and employ chemical suppression technique.
  - The grass is unlikely to grow out of this damage.
  - Delayed action can rob yield from future growth

### Mitigation Strategies

#### Chemical suppression technique requiring 2 applications:

- 1<sup>st</sup> App: 7-10 d after cutting
  - Apply a labeled rate of an insecticide
    - Pyrethroids, such as Baythroid, Karate, Mustang Max, etc. (\$)
    - Spinosad, such as Tracer or Blackhawk (\$ \$\$\$)
- 2<sup>nd</sup> App: repeat 7-10 d later (or 14-20 d after cutting)

### Mitigation Strategies

#### Chemical suppression technique requiring 2 applications:

- Total cost of both applications:
  - ~\$2-3 of product/acre + ~\$4-5 application cost/acre x 2
  - \$12-18/acre/clipping**
- When to apply?
- See signs of the fly – then apply

### Future Research

- Life cycle and reproductive potential of BSM
  - Necessary to refine timing of suppression techniques
- Determine most cost-effective strategies for managing and/or controlling BSM
- Develop a BSM network among the southern states using smartphone apps to report damage of presence of flies – For an early warning systems

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<http://bit.ly/BSM2014>

WHAT WE HAVE LEARNED ABOUT THE BERMUDAGRASS STEM MAGGOT<sup>1</sup>

D.W. Hancock, W.G. Hobson, L.L. Hovine, and E.T. McCullum<sup>2</sup>

Abstract

Since first being discovered in southern Georgia in July 2010, the bermudagrass stem maggot (BSM; *Atherigona reevesae* Villeneuve) has infested and damaged forage bermudagrass (*Cynodon dactylon*) throughout the southeastern United States. Our objectives for this presentation were to summarize the available literature on this new, invasive species and provide additional insight from what is currently known about *Atherigona* spp. The BSM, along with other *Atherigona* spp. are small, muscoid flies native to Central and Southeast Asia. The adult fly of the BSM lays its eggs on bermudagrass leaves. Upon hatching, the BSM larvae slip into the sheath, down the tiller, and penetrate the pseudostem at the first node. The BSM larvae then feeds on the vascular tissue, sap, and (presumably) the subsequent decaying plant material before exiting the tiller, dropping in the soil, and overwintering as a 3<sup>rd</sup> instar. As a result of the second instar, bermudagrass culms experience the necrosis of the terminal tiller on the affected sheath. The affected leaves are easily pulled out of the sheath and show obvious damage near the affected node. In some instances, over 80% of the tillers in a given area may be affected. There is a paucity of information about the lifecycle of *A. reevesae* and how it can be managed or controlled; no serious information is available on basic larva behavior, fly physiology, and the potential differences in resistance among some bermudagrass varieties. Additional research is underway to better understand the lifecycle of this species, confirm and quantify the degree of preference *A. reevesae* has for the different bermudagrass varieties, and quantify the severity of damage to yield, quality, and aesthetics.

Introduction. In the summer of 2010, bermudagrass hay producers in Jeff Davis, Irwin, Pemis, and Elberton in Georgia began receiving a “warning” of their hay fields, generating damage similar to that of severe drought or frost damaged bermudagrass (Hancock, 2012; Fig. 1A). The warning was the result of chlorosis and necrosis in the top two to three leaves of the plant (Fig. 1B). The damaged leaves could easily be pulled from the sheath and the end inside the sheath either showed evidence of insect damage or obvious sheath (Fig. 1C). The damage was clearly not of abiotic stress but rather a consequence of larval feeding. Under controlled conditions, cultured larvae were grown out and allowed to pupate and mature. The resulting adults were subsequently identified as *Atherigona reevesae* Villeneuve, now commonly known as the bermudagrass stem maggot (BSM).

<sup>1</sup> Paper presented at the 2014 American Forage and Grassland Council’s Annual Meeting, January 23-24, Memphis, TN.

<sup>2</sup> Associate Professor and Forage Extension Specialist, Dept. of Crop and Soil Sciences, Professor of Entomology, Dept. of Entomology, Graduate Research Assistant, Dept. of Crop and Soil Sciences, and Graduate Research Assistant, Dept. of Entomology, respectively of the Univ. of Georgia, Athens, GA 30602.

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USDA-ARS Tifton



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